

# No. 356-Egg Freezing for Age-Related Fertility Decline

This Clinical Practice Guideline has been prepared by the Canadian Fertility and Andrology Society (CFAS) Clinical Practice Guideline committee in collaboration with the SOGC Reproductive, Endocrinology and Infertility (REI) committee and reviewed by the SOGC Guideline Management and Oversight Committees and approved by the Boards of the SOGC and CFAS.

Julio Saumet, MD, ART Center, CHU Sainte-Justine Hospital, Montréal, QC  
 Angel Petropanagos, PhD, Dalhousie University, Halifax, NS  
 Karen Buzaglo, MD, Clinique OVO, Montréal, QC  
 Eileen McMahon, RN (EC), NP, Mount Sinai Fertility, Toronto, ON  
 Gunwant Warraich, MD, Olive Fertility Centre, Vancouver, BC  
 Neal Mahutte, MD, The Montréal Fertility Centre, Montréal, QC

**Canadian fertility and andrology society clinical practice guideline committee:** Neal Mahutte, MD (Chair), Montréal, QC; William Buckett, MD, Montréal, QC; Jon Havelock, MD, Vancouver, BC; Kim Liu, MD, Toronto, ON; Jason Min, MD, Calgary, AB; Jeff Roberts, MD, Vancouver, BC; Heather Shapiro, MD, Toronto, ON; Sony Sierra, MD, Toronto, ON; Camille Sylvestre, MD, Montréal, QC. **Reproductive endocrinology and infertility committee:** Anthony P. Cheung, MD (co-chair), Vancouver, BC; Sony Sierra (co-chair), MD, Toronto, ON; Belina Carranza-Mamane, MD, Sherbrooke, QC; Catherine Dwyer, RN, Toronto, ON; James Graham, MD, Victoria, BC; Sarah Healey, MD, St. John's, NL; Robert Hemmings, MD, Westmount, QC;

Tarek Motan, MD, Edmonton, AB; David Smithson, MD, Edmonton, AB; Tannys D.R. Vause, MD, Ottawa, ON; Marta Wais, MD, Toronto, ON; Benjamin Chee-Man Wong, MD, Calgary, AB; Bonnie Woolnough, MD, Edmonton, AB.

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**Key Words:** Oocyte vitrification, social egg freezing, elective egg freezing, fertility preservation, delayed child-bearing

Corresponding Author: Dr. Neal Mahutte, The Montréal Fertility Centre, Montréal, QC. [nealmanhutte@hotmail.com](mailto:nealmanhutte@hotmail.com)

## Abstract

**Objective:** To provide a comprehensive review and evidence based recommendations for Canadian fertility centres that offer social egg freezing.

**Outcomes:** In social egg freezing cycles we evaluated thawed oocyte survival rates, fertilization rates, embryo quality, pregnancy rates, and live birth rates. We also review how these outcomes are impacted by age, ovarian reserve, and the number of eggs cryopreserved. Finally, we discuss the risks of social egg freezing, the alternatives, the critical elements for counselling and informed consent, and future reporting of egg freezing outcome data.

**Evidence:** Published literature was reviewed through searches of MEDLINE and CINAHL using appropriate vocabulary and using key words (“oocyte cryopreservation,” “egg freezing,” “egg vitrification,” “social egg freezing,” and “elective egg freezing”). Results included systematic reviews, randomized controlled trials, controlled clinical trials, and observational studies. Expert opinion based on clinical experience, descriptive studies, or reports of expert committees was also included to discuss aspects of egg freezing not currently rigorously studied.

**Values:** The evidence obtained was reviewed and evaluated by the Clinical Practice Guideline (CPG) Committees of the Canadian Fertility and Andrology Society (CFAS) under the leadership of the principal authors.

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Women have the right and responsibility to make informed decisions about their care in partnership with their health care providers. To facilitate informed choice women should be provided with information and support that is evidence based, culturally appropriate and tailored to their needs. The values, beliefs and individual needs of each woman and her family should be sought and the final decision about the care and treatment options chosen by the woman should be respected.

**Benefits, Harms, and Costs:** Implementation of this guideline should assist the clinician to develop an optimal approach in providing counselling for egg freezing while minimizing harm and improving patient outcomes during treatment.

**Validation:** These guidelines have been reviewed and approved by the membership of the CFAS and by the CPG Committees of CFAS and The Society of Obstetricians and Gynaecologists of Canada (SOGC).

**Sponsors:** CFAS and SOGC.

**Recommendations:**

1. Patients should be advised that thawed oocyte survival rates vary, typically between 80% and 90% (Strong, High).
2. Thawed oocytes should be fertilized using intra cytoplasmic sperm injection and patients should be advised that fertilization rates vary, typically between 70% and 80% (Strong, High).
3. Patients should be advised that vitrified oocytes yield fewer blastocysts than fresh oocytes do (Weak, Moderate).
4. Patients should be advised that there are very limited data on live birth rates after social egg freezing, but that the existing data suggest similar clinical pregnancy rates after transfer of embryos obtained by either vitrified or fresh oocytes (Strong, Moderate).
5. Women considering social egg freezing should be advised that the age at which they freeze their eggs and the number of eggs that are frozen impact the probability that these eggs will enhance their fertility (Strong, Moderate).
6. Ovarian reserve testing should be offered to help predict the number of retrievable eggs from a controlled ovarian stimulation cycle and to properly counsel those women at risk of very low oocyte yield (Strong, High).
7. Women considering social egg freezing should be advised that more than one cycle may be required to obtain the number of mature eggs that is desired (Strong, High).
8. Patients considering social egg freezing should be informed about the risks of controlled ovarian stimulation, oocyte retrieval, and pregnancy at a more advanced maternal age (Strong, Moderate).
9. Patients considering social egg freezing should be advised that there is a chance they may not need to use their frozen eggs and that no guarantees can be made that their frozen eggs would produce a viable pregnancy (Strong, High).
10. Women considering social egg freezing should be counselled about the alternative options (Strong, Moderate).
11. Women undergoing social egg freezing should receive sufficient information to provide informed consent (Strong, High).
12. In vitro fertilization centres offering social egg freezing should provide their patients with an estimate of their chances of success. This estimate should not only consider the published medical literature but also should take into account national data regarding social egg freezing and clinic-specific data regarding cumulative live birth rates per oocyte retrieval (Strong, Low).

## INTRODUCTION

For the past four decades, industrialized countries have experienced an increase in child-bearing age. In Canada, the average age at which women have their first child increased from 23.7 in 1970 to 28.5 in 2011, and more than half of all births now occur in women age 30 and older.<sup>1</sup> The growing delay in parenting has been attributed to improved methods of contraception, as well as economic, professional, educational, and personal changes in modern society.<sup>2–6</sup> In consequence, postponement of parenthood has increased the probability that women will reach an age at which the quantity and quality of their remaining oocytes prevent spontaneous conception if such a conception is desired. It has been estimated that the risk of infertility is approximately 6% at age 20 to 24, 16% at age 30 to 34, and 64% at age 40 to 44.<sup>7</sup> The physiologic decrease in the chances of conception associated with ageing is called age-related fertility decline.

As a result, many women are consulting fertility centres to guard against age-related fertility decline or to optimize future conception. Options include trying to conceive at a younger age, donor sperm insemination, donor egg/embryos, or cryopreserving one's own oocytes for use in future attempts at genetic reproduction.

The first live births after oocyte cryopreservation (egg freezing) occurred over 30 years ago.<sup>8,9</sup> However, due to technical challenges, it was not until recently that the European Society for Human Reproduction and Embryology,<sup>10</sup> the ASRM,<sup>11</sup> and the Canadian Fertility and Andrology Society withdrew their experimental designations for oocyte cryopreservation. Although not intended as an endorsement of elective oocyte cryopreservation, these decisions facilitated consideration of egg freezing as a reproductive option for women who wish to guard against the natural age-related decline in their fertility—so-called “social egg freezing.”

## ABBREVIATIONS

AFC	antral follicle count
AMH	anti-Müllerian hormone
ART	assisted reproductive technology
ASRM	American Society for Reproductive Medicine
CARTR-BORN	Canadian Assisted Reproductive Technologies Register – Better Outcomes Registry & Network
CS	Caesarian section
FSH	follicle-stimulating hormone
ICSI	intracytoplasmic sperm injection
IVF	in vitro fertilization
LBR	Live Birth Rate
RCT	randomized controlled trial

In this guideline, we distinguish social egg freezing from other reasons for oocyte cryopreservation such as pre-gonadotoxic therapy, the unexpected absence of sperm on the day of oocyte retrieval, pre-gender confirmation treatment, or egg freezing performed at the time of IVF because of moral, religious, or legal constraints related to the creation or freezing of supernumerary embryos. Although some have criticized the use of the term “social egg freezing” and recommend instead the term “anticipated gamete exhaustion banking,” social egg freezing remains the most commonly used term for this specific indication.<sup>12</sup>

A substantial number of IVF centres in Canada offer social egg freezing.<sup>13</sup> However, this service is not without controversy.<sup>14</sup> Therefore, the purpose of this guideline is to provide a comprehensive review and evidence-based recommendations for Canadian fertility centres that offer social egg freezing. The quality of evidence was rated using the criteria described in the Grading of Recommendations Assessment, Development, and Evaluation (GRADE) methodology framework, and the interpretation of strong and weak recommendations is described in [Table 1](#) and [Table 2](#).<sup>15</sup>

## WHAT IS THE EFFICACY OF EGG FREEZING?

Oocyte cryopreservation has had to overcome numerous technical challenges related to large cell size, high water content, the delicate chromosomal spindle arrangement of the mature oocyte (an oocyte that has completed the first meiotic division, that has reached metaphase II, and that is capable of being fertilized),<sup>16–19</sup> and the hardening of the zona pellucida that impairs fertilization.<sup>20,21</sup> For many years, the standard procedure for oocyte cryopreservation was slow-freezing. This approach has now largely been replaced by vitrification which uses high initial concentrations of cryoprotectant and ultra-rapid cooling to solidify the cell into a glass-like state without formation of ice crystals.<sup>22</sup> Oocyte vitrification induces less damage to internal structures<sup>23</sup> than slow-freezing and results in superior post-thaw success rates.<sup>24–28</sup> Furthermore, subsequent use of ICSI overcomes fertilization issues related to hardening of the zona pellucida.<sup>29–31</sup>

There is considerable literature regarding the efficacy of oocyte cryopreservation for egg donation, but very limited information on social egg freezing. Indeed, most of the data on social egg freezing come from high-volume centres, and in some cases, the same data overlap different publications. It therefore remains unclear to what extent individual IVF programs can apply the existing data to accurately estimate individual success rates.

**Table 1. Key to grading of recommendations, assessment, development, and evaluation (GRADE)<sup>1</sup>**

Strength of the recommendation	Definition
Strong	Highly confident of the balance between desirable and undesirable consequences (i.e., desirable consequences outweigh the undesirable consequences; or undesirable consequences outweigh the desirable consequences).
Weak <sup>a</sup>	Less confident of the balance between desirable and undesirable consequences.
Quality level of a body of evidence	Definition
High ++++	We are very confident that the true effect lies close to that of the estimate of the effect.
Moderate +++0	We are moderately confident in the effect estimate. The true effect is likely to be close to the estimate of the effect, but there is a possibility that it is substantially different.
Low ++00	Our confidence in the effect estimate is limited. The true effect may be substantially different from the estimate of the effect.
Very Low +000	We have very little confidence in the effect estimate. The true effect is likely to be substantially different from the estimate of effect.

<sup>a</sup>Weak recommendations should not be misinterpreted as weak evidence or uncertainty of the recommendation.

Examples:

Strong, Moderate|+++0: Strong Recommendation, Moderate Quality of Evidence.

Weak, Low|++00: Weak Recommendation, Low Quality of Evidence.

## OOCYTE SURVIVAL RATES

Several studies have analyzed the survival rates of mature vitrified oocytes. The largest experience with egg freezing thus far is derived from egg donation cycles where published survival rates range from 86% to 97%.<sup>32-35</sup> Others have

evaluated survival rates among infertile women, reporting values between 80% and 97%.<sup>36-38</sup> A meta-analysis that included all studies up to June 2013 reported a mean oocyte survival rate of 90% with donor eggs and 86% with non-donor eggs<sup>39</sup> (Table 3). Data on total failure of oocyte survival are sparse and insufficient to make a reliable estimate.<sup>40</sup>

To date, very few publications have evaluated oocyte survival rates, specifically in women undergoing social egg freezing. Cobo et al. described 120 women who returned for fertility treatment and reported a survival rate of 81% of 1080 frozen-thawed oocytes.<sup>32</sup> Doyle et al. described 128 thaw cycles, of which 52 were due to sperm unavailability, 44 for elective limited oocyte fertilization, and 32 for elective egg freezing as the original indications for oocyte freezing, and reported an oocyte survival rate of 86% (Table 3).<sup>41</sup> In comparison, vitrified embryo survival rates typically exceed 90%.<sup>42-44</sup>

**Table 2. Judgement and interpretation of strong and conditional recommendations<sup>15</sup>**

Judgement/ Interpretation	Strong recommendation “We recommend...”	Conditional recommendation “We suggest...”
Judgement by guideline panel	It is clear to the panel that the net desirable consequences of a strategy outweighed the consequences of the alternative strategy.	It is less clear to the panel whether the net desirable consequences of a strategy outweighed the alternative strategy.
Implications for patients	Most individuals in this situation would want the recommended course of action, and only a small proportion would not.	Most individuals in this situation would want the suggested course of action, but many would not.
Implications for clinicians	Most individuals should receive the intervention. Adherence to this recommendation according to the guideline could be used as a quality criterion or performance indicator.	Clinicians should recognize that different choices will be appropriate for each individual and that clinicians must help each individual to arrive at a management decision consistent with his or her values and preferences.
Implications for policy makers	The recommendation can be adopted as policy in most situations.	Policy making will require substantial debate and involvement of various stakeholders.

## FERTILIZATION RATES

Fertilization rates of thawed vitrified oocytes after ICSI in different ART populations have ranged from 71% to 76% with donor eggs,<sup>33,34,40</sup> from 79% to 85% in infertile women,<sup>36-38</sup> and 70% in vitrified eggs from a mixed population of women<sup>41</sup> who underwent egg freezing for various indications (Table 3). In comparison, fertilization rates of fresh oocytes varied from 71% to 82% in donor oocytes cycles (an oocyte donated to a known or anonymous recipient for the purposes of achieving a pregnancy for the intended parent[s]),<sup>33,34</sup> and from 75% to 88% for infertile patients.<sup>36-38</sup>

**Table 3. Oocyte performance after vitrification**

	Number of eggs	Age	Survival rate	Fertilization rate	Implantation rate	Clinical pregnancy rate	Miscarriage rate	Live birth rate
<b>Donor cycles</b>								
Cobo et al. RCT <sup>33</sup>	n = 231	26.7 ± 3.6	97%	76%	41%	N/A	20%	N/A
Cobo et al. RCT <sup>34</sup>	n = 3286	26.7 ± 3.9	93%	74%	40%	55%	N/A	N/A
Solé et al. observational <sup>35</sup>	n = 98	26.1 ± 4.3	86%	78%	34%	54%	21%	43% <sup>b</sup>
<b>Infertile autologous</b>								
Rienzi et al. RCT <sup>36</sup>	n = 24	35.5 ± 4.8	97%	79%	20%	39%	20%	N/A
Parmegiani et al. RCT <sup>37</sup>	n = 168	35 ± 0.8	90%	85%	17%	36%	18%	23% <sup>b</sup>
Chang et al. RCT <sup>38</sup>	n = 86	35.4 ± 2.9	80%	84%	30%	45%	15%	50%
Doyle et al. observational <sup>a,41</sup>	n = 1283	34.9	86%	70%	41%	54%	29%	39%
<b>Social egg freezing</b>								
Cobo et al. observational <sup>32</sup>	n = 1080	37.2	81%	N/A	31%	41%	N/A	21% <sup>b,c</sup>

<sup>a</sup>Included a subset of elective fertility preservation.

<sup>b</sup>Live birth rate per cycle.

<sup>c</sup>Data based on LBR by the time of publication (24 live birth and 26 On-going pregnancy rate (OPR) of 102 transfers).

### **EMBRYO QUALITY**

Some evidence suggests that the embryo quality from thawed vitrified oocytes does not differ from that of fresh oocytes. However, there is heterogeneity in the literature because some centres prefer to transfer at the cleavage stage whereas others do so at the blastocyst stage. A meta-analysis published in 2014 reported an embryo cleavage rate of 92% in fertilized thawed donor eggs and a rate of 84% in fertilized thawed non-donor eggs.<sup>39</sup> The effect of oocyte vitrification on embryo quality has also been evaluated in three RCTs involving patients undergoing treatment for infertility.<sup>36–38</sup> Again, no differences in the quality or percentage of cleavage stage embryos from fresh versus frozen oocytes were found.

### **NUMBER OF BLASTOCYSTS AVAILABLE**

The number of blastocysts available after oocyte thawing has been evaluated by several trials and has yielded conflicting results. In an RCT comparing fresh versus vitrified donor eggs, there were nearly double the number of blastocysts derived from fresh mature oocytes (42/219; 19%) compared to vitrified mature oocytes (24/231; 10%).<sup>33</sup> The same authors conducted another RCT that demonstrated a statistically significant difference in the number of blastocysts comparing fresh versus thawed vitrified donor oocytes (2.5 ± 2.3 vs. 2.0 ± 2.1).<sup>34</sup> In contrast, no differences in the number of day 3 embryos or blastocyst were observed in two RCTs involving infertile patients.<sup>36,38</sup> However, Doyle et al. reported a statistically significant difference in the

number of blastocysts (fresh 66% vs. frozen 51%) in an observational cohort of social and medical egg freezing patients.<sup>41</sup>

### **PREGNANCY RATES**

An RCT with more than 600 patients undergoing vitrified versus fresh egg donation cycles reported results after transfer of a mean of 1.7 embryos. The implantation rate (40%), clinical pregnancy rate (55%), and ongoing pregnancy rate (49%) from frozen eggs was not different compared to fresh donor eggs (Table 3).<sup>34</sup>

Similarly, studies in infertile women report non-significant differences between fresh and frozen eggs. Rienzi et al. randomized 80 women with a mean age of 35 who had at least six eggs at retrieval to either fresh embryo transfer or oocyte vitrification with subsequent thaw/ICSI/transfer.<sup>36</sup> They reported a clinical pregnancy rate of 38.5% and an ongoing pregnancy rate of 30.8% with vitrified eggs compared to 43.2% and 38.8%, respectively, with fresh eggs. A retrospective cohort study involving 1283 thawed vitrified oocytes from a mixed population of egg freezing patients was compared to 243 892 oocytes from fresh IVF.<sup>41</sup> The authors reported no statistical difference in implantation rates (43% vs. 35%) or clinical pregnancy rates (57% vs. 44%) with frozen versus fresh eggs, respectively. However, they also reported a higher pregnancy loss rate per clinical pregnancy with frozen (30%) versus fresh eggs (19%; *P* = 0.048) (Table 3).

## **LIVE BIRTH RATES**

At present, the data regarding live birth rates from social egg freezing cycles are quite limited. Most of the evidence, instead, comes from a varied array of infertility and donor egg cycles, and some of these studies have developed predictive models for the estimation of live birth rate in regards of oocyte number and maternal age.

One of the largest egg donation programs reported 2102 live births from 32 460 thawed donor oocytes. In this program, a mean of 11 donor oocytes allocated per recipient resulted in a live birth rate of 39% per donation cycle. It is important to highlight that 70% of the original stimulated cycles resulted in supernumerary embryos for vitrification and that 632 of these births came from frozen embryos. In fact, the birth rates for the first, second, and third supernumerary frozen-thawed embryo transfers were 32%, 25%, and 24%, respectively. The authors found that the live birth rate per thawed oocyte was 6.5% and the mean number of mature frozen donor oocytes required to achieve a live birth was 15.<sup>40</sup>

Other authors have investigated the live birth rate probabilities in infertile women who underwent oocyte vitrification. One meta-analysis reported a live birth rate of 6% per thawed oocyte.<sup>39</sup> Another meta-analysis estimated that the most discriminatory age for success of oocyte cryopreservation (as measured by live birth) was 36 years (AUC 0.72).<sup>45</sup> In this meta-analysis, the reported live birth rate per transfer of  $2.8 \pm 1.3$  cleavage stage embryos ranged from 12% to 26%. They estimate that thawing up to six oocytes would have a probability of live birth of 24% at 30, 17% at age 36, and 13% at age 40 year (Table 4).<sup>45</sup>

Data regarding live birth rates per frozen egg among women who underwent social egg freezing are quite preliminary. According to Doyle et al., to have a 70% chance of conceiving a child, a woman would need to freeze 14, 15, and 25 eggs at ages 30–34, 35–37, and 38–40, respectively.<sup>41</sup> In their model, it is estimated that the live birth rate per oocyte is 7.4%, 7%, 6.5%, and 5.2% at age <30, age 30–34, age 35–37, and age  $\geq 38$ , respectively. However, the actual live birth rate was 38.6% per transfer cycle in patients with a mean age of 34.9 years. Another recent publication analyzing the outcomes exclusively from social egg freezing (137 women, 1182 thawed oocytes) concluded that there is a statistically significant difference in the live birth rate between women age  $\leq 35$  (50%,  $n = 32$ ) and women older than 35 (22.9%,  $n = 105$ ).<sup>32</sup> Using Kaplan-Meier curves, this group estimated that in social egg freezing patients  $\leq 35$  years, the cumulative live birth rate per 5, 8, and 10 frozen-thawed oocytes would be 15.4%, 40.8%, and 60.5%, respectively,

whereas in women over age 35, it would be 5.1%, 19.9%, and 29.7%, respectively (Table 4).

## **Recommendations**

1. Patients should be advised that thawed oocyte survival rates vary, typically between 80% and 90% (Strong, High).
2. Thawed oocytes should be fertilized using intra cytoplasmic sperm injection, and patients should be advised that fertilization rates vary, typically between 70% and 80% (Strong, High).
3. Patients should be advised that vitrified oocytes yield fewer blastocysts than fresh oocytes do (Weak, Moderate).
4. Patients should be advised that there are very limited data on live birth rates after social egg freezing, but that the existing data suggest similar clinical pregnancy rates after transfer of embryos obtained by either vitrified or fresh oocytes (Strong, Moderate).

## **WHAT FACTORS SHOULD BE ADDRESSED BEFORE CONSIDERING EGG FREEZING? IS THERE A RECOMMENDED AGE?**

The highest probability of live birth in egg freezing programs has been obtained when oocyte cryopreservation is performed before the age of 36.<sup>32,41,45</sup> It is well known that natural fecundity rates decrease after the age of 30, with a steeper decline after the age of 35.<sup>6,7</sup> However, in cost-effectiveness modeling scenarios, the highest live birth rates are attained by egg freezing before the age of 30, but the greatest cost benefit is seen later—that is, for women who freeze their eggs at age 35–37.<sup>46–49</sup> This is not unexpected because women in their 20s or early 30s have relatively stable fertility rates, and the marginal benefit of freezing eggs becomes magnified for these women only if the time between the age of freezing and the age of thawing is significant. Indeed, in the largest social egg freezing series published to date, more than 50% of cases were done at ages 36–39.<sup>32</sup> Despite this late age, only 10% of patients had returned to use their eggs over the time period studied, and the mean storage time was 2.2 years.

## **OVARIAN RESERVE**

Ovarian reserve refers to the number of follicles that may respond to gonadotropin stimulation. Ovarian reserve testing is used to tailor ovarian stimulation in IVF, but it does not necessarily predict conception. Although both the American College of Obstetrics and Gynecology (ACOG)<sup>50</sup> and ASRM<sup>51</sup> have conceptualized ovarian reserve as an indicator of both oocyte number and quality—ovarian reserve

**Table 4. Vitrified oocytes live birth rate**

	Age	Oocyte number (estimated LBR)	Actual LBR per series	
<b>Donor cycles</b> Cobo et al. <sup>40</sup>	21–35	10 (40% <sup>b</sup> )	37% per warming cycle n = 3467 cycles Age = 25.9	
		12 (54% <sup>b</sup> )		
		15 (68% <sup>b</sup> )		
		20 (81% <sup>b</sup> )		
<b>Infertile autologous</b> Pelin Cil et al. <sup>45</sup>	30	2 (21%)	12% to 37% per transfer n = 303 cycles Age = 34.1 ± 4.7	
		4 (23%)		
		6 (24%)		
	35	2 (16%)		
		4 (17%)		
		6 (18%)		
	40	2 (12%)		
		4 (13%)		
		6 (13%)		
	<b>Chang et al.</b> <sup>38</sup>	30–36	1 (8%)	50% per cycle n = 22 cycles Age = 35.4 ± 2.9
		37–39	1 (3%)	
	<b>Doyle et al.</b> <sup>a,41</sup>	30–34	1 (8%)	39% per transfer cycle n = 128 cycles Age = 34.9
5 (30%)				
10 (55%)				
35–37		1 (7%)		
		5 (30%)		
		10 (50%)		
38–40		1 (5%)		
		5 (20%)		
		10 (35%)		
<b>Social egg freezing</b> Cobo et al. <sup>32</sup>		≤35	5 (15%)	20% per cycle, plus 11 ongoing pregnancies at the time of publication n = 148 cycles Age = 37.2
			10 (61%)	
			15 (85%)	
	≥36	5 (5%)		
		10 (30%)		
		11 (36%)		

<sup>a</sup>Included a subset of elective fertility preservation.

<sup>b</sup>Cumulative live birth rate.

testing is only able to directly measure oocyte quantity.<sup>52–56</sup> Several markers are available, and because no single marker is 100% sensitive or specific to determine ovarian reserve, they are often assessed together to give an overall estimate of the number of oocytes that may be retrieved per stimulated cycle.

A basal FSH and estradiol level measured at day 2 to 5 of the menstrual cycle has traditionally been used as a marker of ovarian reserve and to confirm menopause. The main limiting factor of FSH as an ovarian reserve marker is that

it varies from cycle to cycle. However, if basal FSH values are consistently elevated, a poor reproductive prognosis is expected.<sup>57,58</sup>

The AFC is a better prognostic indicator of ovarian stimulation than the basal FSH.<sup>59–62</sup> AFCs decline with age,<sup>63</sup> and lower than expected numbers of antral follicles are a sign of ovarian aging<sup>64</sup> and predict fewer retrievable eggs.<sup>65,66</sup> Although traditionally measured around menstrual cycle day 3, studies show that AFCs remain relatively stable and retain predictive value no matter when they are measured in the cycle.<sup>67,68</sup> A limiting factor is the possibility of inter-observer measurement variability.

As opposed to FSH and AFC, AMH has little inter-cycle variability and may be a better marker for assessing the age-related decline of the ovarian follicular pool and the potential for a poor response to ovarian stimulation.<sup>53,69–76</sup> As the number of ovarian follicles decreases with age, a concomitant decrease in AMH level occurs, which reflects this age-related oocyte depletion.<sup>77</sup> Low AMH levels strongly predict poor response in controlled ovarian stimulation and may be used to help adjust the dose of gonadotropin medication.<sup>78,79</sup>

AMH can be measured at any time throughout the menstrual cycle, as there is only slight inter- and intra-cycle variation in its levels.<sup>80,81</sup> One limitation of AMH testing is the variability of results between the available assays and the inability to compare AMH levels when different assays are used.<sup>82</sup> Another challenge specific to the egg freezing population is the potential for oral contraceptives to suppress by as much as 20% AFCs and AMH levels.<sup>83–85</sup>

**NUMBER OF CYCLES NEEDED**

According to the two groups with the largest experience on social egg freezing, the risk of cycle cancellation due to lack of eggs for vitrification or no transferable embryos range from 1.5% to 4.7%.<sup>32,41</sup> However, another group reported that as many as 21% of women undergoing social egg freezing had ≤3 oocytes at retrieval or had their cycle cancelled because of lack of response to conventional ovarian stimulation.<sup>86</sup> It has been suggested that at least 8–10 vitrified eggs are required to have a reasonable chance of pregnancy and that most women require more than one stimulation cycle to achieve this objective.<sup>32</sup> A more recent model suggested that women age 34 would need to freeze 10 eggs to have a 75% likelihood of having at least one live birth, but that women age 37 would need to freeze 20 eggs and women age 42 would need to freeze 61 eggs to achieve this same probability.<sup>87</sup>

### Recommendations

5. Women considering social egg freezing should be advised that the age at which they freeze their eggs and the number of eggs that are frozen impact the probability that these eggs will enhance their fertility (Strong, Moderate).
6. Ovarian reserve testing should be offered to help predict the number of retrievable eggs from a controlled ovarian stimulation cycle and to properly counsel those women at risk of very low oocyte yield (Strong, High).
7. Women considering social egg freezing should be advised that more than one cycle may be required to obtain the number of mature eggs that is desired (Strong, High).

### WHAT ARE THE RISKS OF SOCIAL EGG FREEZING?

Ovarian hyperstimulation syndrome is a potential consequence of ovarian stimulation. The mild form (bloating, abdominal discomfort) is seen in 20% to 30% of women undergoing controlled ovarian stimulation for IVF, while the most severe form occurs in 1%. Several strategies may be used to prevent ovarian hyperstimulation syndrome in women undergoing social egg freezing, including identifying those patients at highest risk, individualizing gonadotropin dosing, utilizing an antagonist protocol with agonist trigger, and using dopamine agonists.<sup>88–90</sup> The risks related to the procedure of egg retrieval itself include bleeding, infection, injury to intra-peritoneal structures, and conscious sedation/anaesthesia. There is also a potential risk of ovarian torsion from enlarged ovaries, but this risk is extremely rare.<sup>91–93</sup>

The risks of delaying child-bearing via social egg freezing include the risks of pregnancy at a more advanced maternal age.<sup>94,95</sup> Studies in pregnant women age 35 and older demonstrate increased risk for hypertension, gestational diabetes, placenta previa, intrauterine growth restriction, preterm birth, and CS.<sup>96–101</sup> Conversely, the risk of chromosomal abnormalities in the embryo are directly related to the age of the eggs at the time of cryopreservation, and thus, may be reduced in women using their own “younger” eggs. Thus far, there does not appear to be any increased risk of congenital abnormalities related to oocyte vitrification.<sup>102</sup> However, it is important to highlight that long-term data concerning child health and well-being are not yet available.

Other concerns with social egg freezing are related to never having to use the frozen oocytes, which would confer unnecessary financial cost. This is more likely when the women are younger; for example, less than 32 years of age.<sup>103</sup> Indeed,

in the largest series reported to date, only 9% of women who underwent social egg freezing had actually returned to use their eggs (mean age of women who returned 39.2; mean storage time 2.1 years).<sup>32</sup> There is also the possibility of basing future decisions/behaviour on the assumption that one’s frozen eggs will ensure future fertility. Clearly, those eggs may or may not survive the thaw, they may or may not fertilize, and they may or may not produce viable embryos.

### Recommendations

8. Patients considering social egg freezing should be informed about the risks of controlled ovarian stimulation, oocyte retrieval, and pregnancy at a more advanced maternal age (Strong, Moderate).
9. Patients considering social egg freezing should be advised that there is a chance they may not need to use their frozen eggs and that no guarantees can be made that their frozen eggs would produce a viable pregnancy (Strong, High).

### WHAT ARE THE ALTERNATIVES TO SOCIAL EGG FREEZING?

Alternatives to oocyte cryopreservation may involve delayed attempts at pregnancy with or without the use of assisted reproduction, child-bearing at an earlier reproductive age, the pursuit of social non-biological parenthood (through adoption, foster parenting, egg donation), and not having children.

Some women may choose to cryopreserve embryos for future use. Advantages of embryo cryopreservation include fertility centres’ longstanding experience freezing and thawing embryos and that the creation of embryos provides insight into the reproductive potential of what is being cryopreserved. This may be a desirable option for women who wish to reproduce with a male partner with whom they are in a stable relationship. Others may choose to use sperm donation for the creation of embryos which they will freeze for their own personal use. Some individuals may feel moral discomfort or have religious opposition to the cryopreservation of embryos, and the fate of stored embryos may lead to legal disputes in cases of divorce or separation.<sup>16,104</sup>

Women may also choose not to cryopreserve their oocytes and to simply wait until they are ready to start a family. Of those, some will conceive naturally while others may require reproductive assistance. Those who require reproductive assistance may undergo IVF using their own fresh oocytes, donor oocytes, or a donor embryo. For most individuals, the use of one’s own oocytes is desirable but the chances



of a live birth decrease with age. The use of donor oocytes or donor embryos can increase the chances of a live birth, but does not provide genetic connection.

Another option is to attempt pregnancy at an earlier reproductive age. Women should be advised about lifestyle practices and certain medical conditions that may impact fertility.<sup>105</sup> In many cases, conceiving at a younger age would eliminate the need for assisted reproduction. This option may not be ideal given certain personal, social, and financial circumstances.

Social parenthood—for example, domestic or international adoption, fostering, or parenting their (future) partner’s children—can also be a meaningful family-building option. However, there may be legal, financial, and practical barriers to creating families in these ways.

Finally, some women may choose not to have children or refrain from having additional children in the future if they are already a parent. Not all women who inquire about social egg freezing will be certain that they want children, so for some, the choice to remain childless (or have fewer

children) will allow them to pursue other things that they value.

**Recommendations**

- 10. Women considering social egg freezing should be counselled about the alternative options for future conception (Strong, Moderate).

**WHAT ELEMENTS ARE IMPORTANT FOR COUNSELLING AND INFORMED CONSENT?**

Prior to treatment, women considering social egg freezing should be counselled about the medical, physical, psychological, and financial aspects, and social risks/benefits of this technique and the alternatives listed above. Adequate counselling is essential for supporting the process of informed choice (consent or refusal) for social egg freezing. In 2008, the ASRM outlined the essential elements of informed consent for elective oocyte cryopreservation.<sup>106</sup> These elements have been incorporated into and expanded upon in [Table 5](#).

The demographics of women who undergo social egg freezing have been well described. The majority of these women

**Table 5. Essential elements for informed consent for social egg freezing**

<ol style="list-style-type: none"> <li>1. <b>Ovarian stimulation and oocyte retrieval</b> <ol style="list-style-type: none"> <li>a) The side effects and risks associated with required medications.</li> <li>b) The requirements for blood sampling, transvaginal ultrasound, and oocyte retrieval.</li> <li>c) The potential risks of oocyte retrieval.</li> <li>d) A reasonable estimate of the number of oocytes that may be retrieved.</li> </ol> </li> <li>2. <b>Cryopreservation and storage</b> <ol style="list-style-type: none"> <li>a) The cryopreservation method to be used.</li> <li>b) The location where the oocytes will be stored.</li> <li>c) The possibility that their cryopreserved oocytes will have to be transferred to a different location if the storage facility moves or ceases operation.</li> <li>d) The possibility that cryopreserved oocytes may be lost or damaged.</li> <li>e) The expected thaw-survival rate for cryopreserved oocytes including the possibility that none survive.</li> <li>f) The fact that cryopreservation does not guarantee live birth.</li> </ol> </li> <li>3. <b>IVF</b> <ol style="list-style-type: none"> <li>a) The requirement for IVF-ICSI and the associated risks for the woman and resulting offspring.</li> <li>b) The expected success rates per thawed oocyte for fertilization, embryo development, pregnancy, and live birth (including clinic-specific data if available).</li> <li>c) Age-specific expected outcomes for IVF using cryopreserved vs. fresh oocytes.</li> <li>d) Any applicable requirements for single embryo transfer.</li> </ol> </li> <li>4. <b>Finances</b> <ol style="list-style-type: none"> <li>a) The cost of oocyte cryopreservation (including medications).</li> <li>b) Annual storage fees for cryopreserved oocytes.</li> <li>c) The estimated cost of later IVF-ICSI with embryo transfer and possible embryo cryopreservation.</li> <li>d) Applicable exclusions or limitations on public funding for IVF.</li> </ol> </li> <li>5. <b>Advanced maternal age</b> <ol style="list-style-type: none"> <li>a) The maternal and fetal risks associated with childbearing at an advanced age with natural or ART conceived pregnancies, and</li> <li>b) The potential challenges and benefits of parenting at a more advanced reproductive age.</li> </ol> </li> <li>6. <b>Use or disposition of cryopreserved oocytes</b> <ol style="list-style-type: none"> <li>a) That cryopreserved oocytes may be destroyed if storage fees are not paid or if they exceed any clinic-specific time limits for the use of stored materials or access to IVF, or in the event of their death.</li> <li>b) That cryopreserved oocytes that are not used for the patient’s reproduction may be disposed of (at the patient’s request).</li> <li>c) That with prior consent, the cryopreserved oocytes may be donated to another couple or individual for the purposes of reproduction; they may be used for research aimed at improving reproductive techniques or training; or they may be used for other research purposes.</li> <li>d) That Canada currently prohibits the sale of gametes (including oocytes); and</li> <li>e) That there may be moral/ethical implications concerning the future disposition or use of unused cryopreserved oocytes.</li> </ol> </li> </ol>
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**Table 6. Cumulative live birth rate per batch of oocytes retrieved (from CARTR-BORN Jan 1, 2013–Dec 31, 2014, after all resulting embryos, fresh and frozen, have been transferred or discarded)**

Number of mature eggs retrieved	Age of oocyte provider				
	Less than 30	30–34	35–37	38–40	41–42
<5 eggs	20.3%	21.3%	17.0%	11.7%	5.9%
5–9 eggs	35.6%	38.3%	31.4%	23.8%	13.4%
10–14 eggs	46.8%	45.3%	41.9%	31.8%	21.8%
15–19 eggs	46.8%	51.1%	45.5%	33.2%	30.3%
20–24 eggs	47.6%	53.1%	50.3%	45.9%	26.7%
25+ eggs	43.7%	48.9%	53.1%	54.0%	40.9%
Total	38.5%	38.3%	30.9%	21.8%	12.5%

tend to be older than 35, single, childless, heterosexual, and with higher education.<sup>3,32,107</sup> A survey of women post-oocyte cryopreservation showed that almost 100% still desired a pregnancy, around 50% were in a relationship, and one-third were trying to conceive. Of these patients, 17% had tried to conceive naturally in the past 12 months without success and 29% reported a pregnancy/delivery since freezing their eggs. Despite changes in relationship and/or parenting status, when women were asked about regrets from social egg freezing, the overwhelming majority stated that they would do it again but at a younger age.<sup>107</sup>

### Recommendations

11. Women undergoing social egg freezing should receive sufficient information to provide informed consent (Strong, High).

### HOW DO WE REPORT OUTCOME DATA WHEN SO LITTLE DATA ARE AVAILABLE?

Ideally, all clinics that offer social egg freezing would be able to report their clinic-specific outcome data. However, because the number of women undergoing social egg freezing is relatively small and because the number of women who have thus far returned to use their frozen eggs is even smaller, it will take many years, if not decades, for most clinics to have reliable, representative, age-specific data. As a result, many clinics refer to the medical literature and imply that these data represent the chance for success. Unfortunately, the data reported from large international centres may, or may not, have external validity.

An alternative may be to report national data for social egg freezing from the CARTR-BORN registry. Although not centre-specific, this should reflect the scope and diversity of centres that offer social egg freezing in Canada. However, these data have only recently come into the CARTR-BORN registry, and we do not yet have live birth data to report.

Another alternative, or supplement, may be to report cumulative live birth rates per oocyte retrieval. This, too, is a recent addition to CARTR-BORN and provides the option of both national and centre-specific, age-specific data (Table 6). Clearly, one cannot assume that the results with social egg freezing would be equivalent to autologous, newly completed IVF cycles, but this could provide a way of estimating the upper limit for the success rate of this technique.

### Recommendations

12. In vitro fertilization centres offering social egg freezing should provide their patients with an estimate of their chances of success. This estimate not only should consider the published medical literature but also should take into account national data regarding social egg freezing and, if available, clinic-specific data regarding cumulative live birth rates per oocyte retrieval (Strong, Low).

### CONCLUSION

Social egg freezing is an increasingly common method for women to attempt to guard against the natural age-related fertility decline. Although existing data are limited, they demonstrate an acceptable pregnancy rate, as well as some psychosocial benefit. Nevertheless, caution is required when counselling women about their chances of success. As with all medical interventions, a thoughtful discussion about the risks, benefits, and alternatives to social egg freezing is paramount.

### REFERENCES

1. Milan A. Fertility: overview, 2009 to 2011. In: Division D, editor. Statistics Canada. Statistics Canada; 2015. Available at: <http://www.statcan.gc.ca/pub/91-209-x/2013001/article/11784-eng.pdf>. Accessed on March 2, 2017.
2. Mill C, Enders J, Montanaro C, et al. Delayed parenthood on the rise: a call for upstream preconception health promotion in Canada. *Can J Public Health* 2016;107:e333–5.

3. Hodes-Wertz B, Druckenmiller S, Smith M, et al. What do reproductive-age women who undergo oocyte cryopreservation think about the process as a means to preserve fertility? *Fertil Steril* 2013;100:1343–9.
4. Hammarberg K, Clarke VE. Reasons for delaying childbearing—a survey of women aged over 35 years seeking assisted reproductive technology. *Aust Fam Physician* 2005;34:187–8, 206.
5. Mills M, Rindfuss RR, McDonald P, et al. Why do people postpone parenthood? Reasons and social policy incentives. *Hum Reprod Update* 2011;17:848–60.
6. van Noord-Zaadstra BM, Looman CW, Alsbach H, et al. Delaying childbearing: effect of age on fecundity and outcome of pregnancy. *BMJ* 1991;302:1361–5.
7. Menken J, Trussell J, Larsen U. Age and infertility. *Science* 1986;233:1389–94.
8. Chen C. Pregnancy after human oocyte cryopreservation. *Lancet* 1986;1:884–6.
9. van Uem JF, Siebzehrubl ER, Schuh B, et al. Birth after cryopreservation of unfertilized oocytes. *Lancet* 1987;1:752–3.
10. ESHRE Task Force on Ethics and Law, Dondorp W, de Wert G, et al. Oocyte cryopreservation for age-related fertility loss. *Hum Reprod* 2012;27:1231–7.
11. Practice Committees of American Society for Reproductive Medicine; Society for Assisted Reproductive Technology. Mature oocyte cryopreservation: a guideline. *Fertil Steril* 2013;99:37–43.
12. Stoop D, van der Veen F, Deneyer M, et al. Oocyte banking for anticipated gamete exhaustion (AGE) is a preventive intervention, neither social nor nonmedical. *Reprod Biomed Online* 2014;28:548–51.
13. Liu KE, Greenblatt EM. Oocyte cryopreservation in Canada: a survey of Canadian ART clinics. *J Obstet Gynaecol Can* 2012;34:250–6.
14. Petropanagos A, Cattapan A, Baylis F, et al. Social egg freezing: risk, benefits and other considerations. *CMAJ* 2015;187:666–9.
15. GRADE Working Group. Schünemann H, Brožek J, Guyatt G, Oxman A, editors. GRADE handbook. 2013. Available at: <http://gdt.guidelinedevelopment.org/app/handbook/handbook.html>. Accessed on June 8, 2017.
16. Argyle CE, Harper JC, Davies MC. Oocyte cryopreservation: where are we now? *Hum Reprod Update* 2016;22:440–9.
17. Paynter SJ, Cooper A, Gregory L, et al. Permeability characteristics of human oocytes in the presence of the cryoprotectant dimethylsulphoxide. *Hum Reprod* 1999;14:2338–42.
18. Pickering SJ, Braude PR, Johnson MH, et al. Transient cooling to room temperature can cause irreversible disruption of the meiotic spindle in the human oocyte. *Fertil Steril* 1990;54:102–8.
19. Vincent C, Johnson MH. Cooling, cryoprotectants, and the cytoskeleton of the mammalian oocyte. *Oxf Rev Reprod Biol* 1992;14:73–100.
20. Matson PL, Graefling J, Junk SM, et al. Cryopreservation of oocytes and embryos: use of a mouse model to investigate effects upon zona hardness and formulate treatment strategies in an in-vitro fertilization programme. *Hum Reprod* 1997;12:1550–3.
21. Johnson MH, Pickering SJ, George MA. The influence of cooling on the properties of the zona pellucida of the mouse oocyte. *Hum Reprod* 1988;3:383–7.
22. Antinori M, Licata E, Dani G, et al. Cryotop vitrification of human oocytes results in high survival rate and healthy deliveries. *Reprod Biomed Online* 2007;14:72–9.
23. Ciotti PM, Porcu E, Notarangelo L, et al. Meiotic spindle recovery is faster in vitrification of human oocytes compared to slow freezing. *Fertil Steril* 2009;91:2399–407.
24. Oktay K, Cil AP, Bang H. Efficiency of oocyte cryopreservation: a meta-analysis. *Fertil Steril* 2006;86:70–80.
25. Cao YX, Xing Q, Li L, et al. Comparison of survival and embryonic development in human oocytes cryopreserved by slow-freezing and vitrification. *Fertil Steril* 2009;92:1306–11.
26. Fadini R, Brambillasca F, Renzini MM, et al. Human oocyte cryopreservation: comparison between slow and ultrarapid methods. *Reprod Biomed Online* 2009;19:171–80.
27. Smith GD, Serafini PC, Fioravanti J, et al. Prospective randomized comparison of human oocyte cryopreservation with slow-rate freezing or vitrification. *Fertil Steril* 2010;94:2088–95.
28. Cobo A, Diaz C. Clinical application of oocyte vitrification: a systematic review and meta-analysis of randomized controlled trials. *Fertil Steril* 2011;96:277–85.
29. Kazem R, Thompson LA, Srikantharajah A, et al. Cryopreservation of human oocytes and fertilization by two techniques: in-vitro fertilization and intracytoplasmic sperm injection. *Hum Reprod* 1995;10:2650–4.
30. Porcu E, Fabbri R, Seracchioli R, et al. Birth of a healthy female after intracytoplasmic sperm injection of cryopreserved human oocytes. *Fertil Steril* 1997;68:724–6.
31. Fabbri R, Porcu E, Marsella T, et al. Oocyte cryopreservation. *Hum Reprod* 1998;13:98–108.
32. Cobo A, Garcia-Velasco JA, Coello A, et al. Oocyte vitrification as an efficient option for elective fertility preservation. *Fertil Steril* 2016;105:755–64, e8.
33. Cobo A, Kuwayama M, Perez S, et al. Comparison of concomitant outcome achieved with fresh and cryopreserved donor oocytes vitrified by the Cryotop method. *Fertil Steril* 2008;89:1657–64.
34. Cobo A, Meseguer M, Remohi J, et al. Use of cryo-banked oocytes in an ovum donation programme: a prospective, randomized, controlled, clinical trial. *Hum Reprod* 2010;25:2239–46.
35. Sole M, Santalo J, Boada M, et al. How does vitrification affect oocyte viability in oocyte donation cycles? A prospective study to compare outcomes achieved with fresh versus vitrified sibling oocytes. *Hum Reprod* 2013;28:2087–92.
36. Rienzi L, Romano S, Albricci L, et al. Embryo development of fresh “versus” vitrified metaphase II oocytes after ICSI: a prospective randomized sibling-oocyte study. *Hum Reprod* 2010;25:66–73.
37. Parmegiani L, Cognigni GE, Bernardi S, et al. Efficiency of aseptic open vitrification and hermetical cryostorage of human oocytes. *Reprod Biomed Online* 2011;23:505–12.
38. Chang CC, Elliott TA, Wright G, et al. Prospective controlled study to evaluate laboratory and clinical outcomes of oocyte vitrification obtained in in vitro fertilization patients aged 30 to 39 years. *Fertil Steril* 2013;99:1891–7.
39. Potdar N, Gelbaya TA, Nardo LG. Oocyte vitrification in the 21st century and post-warming fertility outcomes: a systematic review and meta-analysis. *Reprod Biomed Online* 2014;29:159–76.
40. Cobo A, Garrido N, Pellicer A, et al. Six years’ experience in ovum donation using vitrified oocytes: report of cumulative outcomes, impact of

- storage time, and development of a predictive model for oocyte survival rate. *Fertil Steril* 2015;104:1426–34, e1–8.
41. Doyle JO, Richter KS, Lim J, et al. Successful elective and medically indicated oocyte vitrification and warming for autologous in vitro fertilization, with predicted birth probabilities for fertility preservation according to number of cryopreserved oocytes and age at retrieval. *Fertil Steril* 2016;105:459–66, e2.
  42. Liebermann J. Vitrification of human blastocysts: an update. *Reprod Biomed Online* 2009;4:4328.
  43. Loutradi KE, Kolibianakis EM, Venetis CA, et al. Cryopreservation of human embryos by vitrification or slow freezing: a systematic review and meta-analysis. *Fertil Steril* 2008;90:186–93.
  44. Balaban B, Urman B, Ata B, et al. A randomized controlled study of human Day 3 embryo cryopreservation by slow freezing or vitrification: vitrification is associated with higher survival, metabolism and blastocyst formation. *Hum Reprod* 2008;23:1976–82.
  45. Cil AP, Bang H, Oktay K. Age-specific probability of live birth with oocyte cryopreservation: an individual patient data meta-analysis. *Fertil Steril* 2013;100:492–9, e3.
  46. Devine K, Mumford SL, Goldman KN, et al. Baby budgeting: oocyte cryopreservation in women delaying reproduction can reduce cost per live birth. *Fertil Steril* 2015;103:1446–53, e1–2.
  47. van Loendersloot LL, Moolenaar LM, Mol BW, et al. Expanding reproductive lifespan: a cost-effectiveness study on oocyte freezing. *Hum Reprod* 2011;26:3054–60.
  48. Hirshfeld-Cytron J, Grobman WA, Milad MP. Fertility preservation for social indications: a cost-based decision analysis. *Fertil Steril* 2012;97:665–70.
  49. Mesen TB, Mersereau JE, Kane JB, et al. Optimal timing for elective egg freezing. *Fertil Steril* 2015;103:1551–6, e1–4.
  50. Committee on Gynecologic Practice. Committee opinion no. 618: ovarian reserve testing. *Obstet Gynecol* 2015;125:268–73.
  51. Practice Committee of the American Society for Reproductive Medicine. Testing and interpreting measures of ovarian reserve: a committee opinion. *Fertil Steril* 2015;103:e9–17.
  52. Melo MA, Garrido N, Alvarez C, et al. Antral follicle count (AFC) can be used in the prediction of ovarian response but cannot predict the oocyte/embryo quality or the in vitro fertilization outcome in an egg donation program. *Fertil Steril* 2009;91:148–56.
  53. McIlveen M, Skull JD, Ledger WL. Evaluation of the utility of multiple endocrine and ultrasound measures of ovarian reserve in the prediction of cycle cancellation in a high-risk IVF population. *Hum Reprod* 2007;22:778–85.
  54. Younis JS, Ben-Ami M, Ben-Shlomo I. The Bologna criteria for poor ovarian response: a contemporary critical appraisal. *J Ovarian Res* 2015;8:76.
  55. Ripley M, Lanes A, Leveille MC, et al. Does ovarian reserve predict egg quality in unstimulated therapeutic donor insemination cycles? *Fertil Steril* 2015;103:1170–5, e2.
  56. Tremellen K, Kolo M. Serum anti-Mullerian hormone is a useful measure of quantitative ovarian reserve but does not predict the chances of live-birth pregnancy. *Aust N Z J Obstet Gynaecol* 2010;50:568–72.
  57. Kwee J, Schats R, McDonnell J, et al. Intercycle variability of ovarian reserve tests: results of a prospective randomized study. *Hum Reprod* 2004;19:590–5.
  58. Roberts JE, Spandorfer S, Fasouliotis SJ, et al. Taking a basal follicle-stimulating hormone history is essential before initiating in vitro fertilization. *Fertil Steril* 2005;83:37–41.
  59. Polyzos NP, Tournaye H, Guzman L, et al. Predictors of ovarian response in women treated with corifollitropin alfa for in vitro fertilization/intracytoplasmic sperm injection. *Fertil Steril* 2013;100:430–7.
  60. Fleming R, Seifer DB, Frattarelli JL, et al. Assessing ovarian response: antral follicle count versus anti-Mullerian hormone. *Reprod Biomed Online* 2015;31:486–96.
  61. Broer SL, van Disseldorp J, Broeze KA, et al. Added value of ovarian reserve testing on patient characteristics in the prediction of ovarian response and ongoing pregnancy: an individual patient data approach. *Hum Reprod Update* 2013;19:26–36.
  62. Jayaprakasan K, Campbell B, Hopkisson J, et al. A prospective, comparative analysis of anti-Mullerian hormone, inhibin-B, and three-dimensional ultrasound determinants of ovarian reserve in the prediction of poor response to controlled ovarian stimulation. *Fertil Steril* 2010;93:855–64.
  63. Bozdag G, Calis P, Zengin D, et al. Age related normogram for antral follicle count in general population and comparison with previous studies. *Eur J Obstet Gynecol Reprod Biol* 2016;206:120–4.
  64. La Marca A, Spada E, Sighinolfi G, et al. Age-specific nomogram for the decline in antral follicle count throughout the reproductive period. *Fertil Steril* 2011;95:684–8.
  65. Gibreel A, Maheshwari A, Bhattacharya S, et al. Ultrasound tests of ovarian reserve; a systematic review of accuracy in predicting fertility outcomes. *Hum Fertil* 2009;12:95–106.
  66. Jayaprakasan K, Chan Y, Islam R, et al. Prediction of in vitro fertilization outcome at different antral follicle count thresholds in a prospective cohort of 1,012 women. *Fertil Steril* 2012;98:657–63.
  67. Rombauts L, Onwude JL, Chew HW, et al. The predictive value of antral follicle count remains unchanged across the menstrual cycle. *Fertil Steril* 2011;96:1514–8.
  68. Mavrelou D, Al Chami A, Talaulikar V, et al. Variation in antral follicle counts at different times in the menstrual cycle: does it matter? *Reprod Biomed Online* 2016;33:174–9.
  69. Ferraretti AP, La Marca A, Fauser BC, et al. ESHRE consensus on the definition of “poor response” to ovarian stimulation for in vitro fertilization: the Bologna criteria. *Hum Reprod* 2011;26:1616–24.
  70. Broer SL, Dolleman M, van Disseldorp J, et al. Prediction of an excessive response in in vitro fertilization from patient characteristics and ovarian reserve tests and comparison in subgroups: an individual patient data meta-analysis. *Fertil Steril* 2013;100:420–9, e7.
  71. Silva JB, Panaino TR, Tamm MA, et al. Prediction of metaphase II oocytes according to different serum Anti-Mullerian hormone (AMH) levels in antagonist ICSI cycles. *JBRA Assist Reprod* 2016;20:222–6.
  72. Iliodromiti S, Anderson RA, Nelson SM. Technical and performance characteristics of anti-Mullerian hormone and antral follicle count as biomarkers of ovarian response. *Hum Reprod Update* 2015;21:698–710.
  73. Ganidou MA, Kolibianakis EM, Venetis CA, et al. Is assessment of anti-Mullerian hormone and/or antral follicle count useful in the prediction of ovarian response in expected normal responders treated with a fixed dose of recombinant FSH and GnRH antagonists? A prospective observational study. *Gynecol Endocrinol* 2014;30:817–21.
  74. Arce JC, La Marca A, Mirner Klein B, et al. Antimullerian hormone in gonadotropin releasing-hormone antagonist cycles: prediction of

- ovarian response and cumulative treatment outcome in good-prognosis patients. *Fertil Steril* 2013;99:1644–53.
75. Kunt C, Ozaksit G, Keskin Kurt R, et al. Anti-Mullerian hormone is a better marker than inhibin B, follicle stimulating hormone, estradiol or antral follicle count in predicting the outcome of in vitro fertilization. *Arch Gynecol Obstet* 2011;283:1415–21.
  76. Nardo LG, Gelbaya TA, Wilkinson H, et al. Circulating basal anti-Mullerian hormone levels as predictor of ovarian response in women undergoing ovarian stimulation for in vitro fertilization. *Fertil Steril* 2009;92:1586–93.
  77. La Marca A, Spada E, Grisendi V, et al. Normal serum anti-Mullerian hormone levels in the general female population and the relationship with reproductive history. *Eur J Obstet Gynecol Reprod Biol* 2012;163:180–4.
  78. La Marca A, Sighinolfi G, Radi D, et al. Anti-Mullerian hormone (AMH) as a predictive marker in assisted reproductive technology (ART). *Hum Reprod Update* 2010;16:113–30.
  79. Heidar Z, Bakhtiyari M, Mirzamoradi M, et al. Prediction of different ovarian responses using anti-Mullerian hormone following a long agonist treatment protocol for IVF. *J Endocrinol Invest* 2015;38:1007–15.
  80. Kissell KA, Danaher MR, Schisterman EF, et al. Biological variability in serum anti-Mullerian hormone throughout the menstrual cycle in ovulatory and sporadic anovulatory cycles in eumenorrheic women. *Hum Reprod* 2014;29:1764–72.
  81. Rezende CP, Rocha AL, Dela Cruz C, et al. Serum antimullerian hormone measurements with second generation assay at two distinct menstrual cycle phases for prediction of cycle cancellation, pregnancy and live birth after in vitro fertilization. *J Assist Reprod Genet* 2014;31:1303–10.
  82. Rustamov O, Smith A, Roberts SA, et al. The measurement of anti-Mullerian hormone: a critical appraisal. *J Clin Endocrinol Metab* 2014;99:723–32.
  83. D'Arpe S, Di Felicianantonio M, Candelieri M, et al. Ovarian function during hormonal contraception assessed by endocrine and sonographic markers: a systematic review. *Reprod Biomed Online* 2016;33:436–48.
  84. Birch Petersen K, Hvidman HW, Forman JL, et al. Ovarian reserve assessment in users of oral contraception seeking fertility advice on their reproductive lifespan. *Hum Reprod* 2015;30:2364–75.
  85. Deb S, Campbell BK, Pincott-Allen C, et al. Quantifying effect of combined oral contraceptive pill on functional ovarian reserve as measured by serum anti-Mullerian hormone and small antral follicle count using three-dimensional ultrasound. *Ultrasound Obstet Gynecol* 2012;39:574–80.
  86. Tsafirir A, Haimov-Kochman R, Margalioth EJ, et al. Ovarian stimulation for oocyte cryopreservation for prevention of age-related fertility loss: one in five is a low responder. *Gynecol Endocrinol* 2015;31:779–82.
  87. Goldman RH, Racowsky C, Farland LV, et al. Predicting the likelihood of live birth for elective oocyte cryopreservation: a counseling tool for physicians and patients. *Hum Reprod* 2017;32:853–9.
  88. Humaidan P, Quartarolo J, Papanikolaou EG. Preventing ovarian hyperstimulation syndrome: guidance for the clinician. *Fertil Steril* 2010;94:389–400.
  89. Nayak SR, Wakim AN. Random-start gonadotropin-releasing hormone (GnRH) antagonist-treated cycles with GnRH agonist trigger for fertility preservation. *Fertil Steril* 2011;96:e51–4.
  90. Oktay K, Turkcuoglu I, Rodriguez-Wallberg KA. GnRH agonist trigger for women with breast cancer undergoing fertility preservation by aromatase inhibitor/FSH stimulation. *Reprod Biomed Online* 2010;20:783–8.
  91. Gelbaya TA. Short and long-term risks to women who conceive through in vitro fertilization. *Hum Fertil* 2010;13:19–27.
  92. Maxwell KN, Cholst IN, Rosenwaks Z. The incidence of both serious and minor complications in young women undergoing oocyte donation. *Fertil Steril* 2008;90:2165–71.
  93. Bodri D, Guillen JJ, Polo A, et al. Complications related to ovarian stimulation and oocyte retrieval in 4052 oocyte donor cycles. *Reprod Biomed Online* 2008;17:237–43.
  94. Biro MA, Davey MA, Carolan M, et al. Advanced maternal age and obstetric morbidity for women giving birth in Victoria, Australia: a population-based study. *Aust N Z J Obstet Gynaecol* 2012;52:229–34.
  95. Lemoine ME, Ravitsky V. Sleepwalking into infertility: the need for a public health approach toward advanced maternal age. *Am J Bioeth* 2015;15:37–48.
  96. Carolan MC, Davey MA, Biro M, et al. Very advanced maternal age and morbidity in Victoria, Australia: a population based study. *BMC Pregnancy Childbirth* 2013;13:80.
  97. Ludford I, Scheil W, Tucker G, Grivell R. Pregnancy outcomes for nulliparous women of advanced maternal age in South Australia, 1998–2008. *Aust N Z J Obstet Gynaecol* 2012;52:235–41.
  98. Cleary-Goldman J, Malone FD, Vidaver J, et al. Impact of maternal age on obstetric outcome. *Obstet Gynecol* 2005;105:983–90.
  99. Ziadeh S, Yahaya A. Pregnancy outcome at age 40 and older. *Arch Gynecol Obstet* 2001;265:30–3.
  100. Gilbert WM, Nesbitt TS, Danielsen B. Childbearing beyond age 40: pregnancy outcome in 24,032 cases. *Obstet Gynecol* 1999;93:9–14.
  101. Jahromi BN, Hussein Z. Pregnancy outcome at maternal age 40 and older. *Taiwan J Obstet Gynecol* 2008;47:318–21.
  102. Noyes N, Porcu E, Borini A. Over 900 oocyte cryopreservation babies born with no apparent increase in congenital anomalies. *Reprod Biomed Online* 2009;18:769–76.
  103. Robertson JA. Egg freezing and egg banking: empowerment and alienation in assisted reproduction. *J Law Biosci* 2014;1:113–36.
  104. Pennings G. What are the ownership rights for gametes and embryos? Advance directives and the disposition of cryopreserved gametes and embryos. *Hum Reprod* 2000;15:979–86.
  105. Practice Committee of the American Society for Reproductive Medicine in collaboration with the Society for Reproductive Endocrinology and Infertility (electronic address: ASRM@asrm.org), Practice Committee of the American Society for Reproductive Medicine in collaboration with the Society for Reproductive Endocrinology and Infertility. Optimizing natural fertility: a committee opinion. *Fertil Steril* 2017;107:52–8.
  106. Practice Committee of Society for Assisted Reproductive Technology; Practice Committee of American Society for Reproductive Medicine. Essential elements of informed consent for elective oocyte cryopreservation: a Practice Committee opinion. *Fertil Steril* 2008;90:S134–5.
  107. Stoop D, Maes E, Polyzos NP, et al. Does oocyte banking for anticipated gamete exhaustion influence future relational and reproductive choices? A follow-up of bankers and non-bankers. *Hum Reprod* 2015;30:338–44.